

# Development of a High-Throughput Molecular Imaging-Based Orthotopic Hepatocellular Carcinoma Model

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## Abstract

We have developed a novel orthotopic rat hepatocellular (HCC) model and have assessed the ability to use bioluminescence imaging (BLI), positron emission tomography (PET), and ultrasound for early tumor detection and monitoring of disease progression.

Briefly, rat HCC cells were stably transfected with click beetle red as a reporter gene for BLI. Tumor cells were injected under direct visualization into the left or middle lobe of the liver in 37 rats. In six animals, serial PET, BLI, and ultrasound imaging were performed at 10-time points in 28 days. The remainder of the animals underwent PET imaging at 14 days. Tumor implantation was successful in 34 of 37 animals (91.9%). In the six animals that underwent serial imaging, tumor formation was first detected with BLI on Day 4 with continued increase through Day 21, and hypermetabolic activity on PET was first noted on Days 14-15 with continued increase through Day 28. PET activity was seen on Day 14 in the 28 other animals that demonstrated tumor development. Anatomic tumor formation was detected with ultrasound at Days 10-12 with continued growth through Day 28. The first metastases were detected by PET after Day 24.

We have successfully developed and validated a novel orthotopic HCC small animal model that permits longitudinal assessment of change in tumor size using molecular imaging techniques. BLI is the most sensitive imaging method for detection of early tumor formation and growth. This model permits high-throughput *in vivo* evaluation of image-guided therapies.

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**Categories:** Radiology, General Surgery, Oncology

**Keywords:** small animal model, hepatocellular carcinoma, molecular imaging, tumor growth, bioluminescence imaging, positron emission tomography

## Introduction

Hepatocellular carcinoma (HCC) is the fifth most common neoplasm in the world, with a worldwide incidence of more than 500,000 new cases diagnosed annually [1]. Despite a broad armamentarium of therapies aimed at treating HCC - including chemotherapeutic medications, percutaneous ablation, transarterial chemoembolization, radioembolization, surgical resection, and transplantation - long-term disease-free survival remains elusive in the vast majority of patients. First-line treatments include resection and liver transplantation [2]. Unfortunately, only 10% of HCC patients are eligible for surgical treatment [3]. Medications, such as sorafenib, offer only modest survival benefits in select patients [4]. Therefore, image-guided interventions, such as transarterial and percutaneous therapies, are the mainstays of therapy for non-surgical candidates.

Historically, research in image-guided interventions has relied on the use of large animals. This was driven by a focus on the development of mechanical medical devices. However, large animal experiments are extremely expensive and time-intensive. Moreover, if the longitudinal effects of a therapy are to be measured, multiple animals must enter the same treatment arm of the experiment, but then must be euthanized at different time points in order to have histologic correlation. This increases the number of animals needed to generate significant results.

As disease treatment shifts from a mechanical to a molecular approach, the important feature of an appropriate animal model is not whether the organ or vessel size matches that of humans but rather whether the model mimics the disease at a molecular and cellular level. Many of the published studies investigating HCC image-guided interventions currently utilize the rabbit VX2 model [5-15]. This model enjoys the benefits of a convenient size for vascular catheterizations and percutaneous ablative therapies. However, it suffers from the shortcomings of large animal models, and it is limited in that it is created with a squamous carcinoma cell line and, hence, does not represent a true hepatocellular carcinoma.

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In designing a model for image-guided molecular interventions for HCC, we sought a few key characteristics. First, the model must be cell-specific for HCC. Second, the model must be small enough for high-throughput studies and yet large enough to test intraparenchymal and intra-arterial delivery routes. The Morris hepatoma model - which uses rat HCC cells from an inbred immune-competent rat line - met both of these criteria [16-20]. Different techniques have been described to implant the hepatoma cells in the rat liver. We optimized the technique, described below, to reproducibly implant a single focus of tumor in the liver while minimizing the spillage of cells into the peritoneal cavity.

A third characteristic we desired in our model was the ability to serially assess the growth and response to therapy of the tumor without euthanizing multiple animals at different time points. Not only would this decrease the time and resources needed to complete the experiment, but it also would also allow us to better appreciate relatively small differences in response to therapy. One difficulty with traditional models that require euthanasia at different time points is that it is impossible to guarantee that the tumor in each subject is the same size at any given time point. In order to overcome this inherent variability, large numbers of animals must be studied to assess effect of therapy, and there is still a risk that the results are skewed by differences in tumor growth in the different study arms. To allow quantitative serial evaluation of the tumor, we stably transfected the tumor cells with a bioluminescence gene prior to implantation. We then assessed our ability to functionally and morphologically characterize the tumor with multiple non-invasive imaging modalities.

## Materials And Methods

### Generation of McA-RH7777-CBRLuc

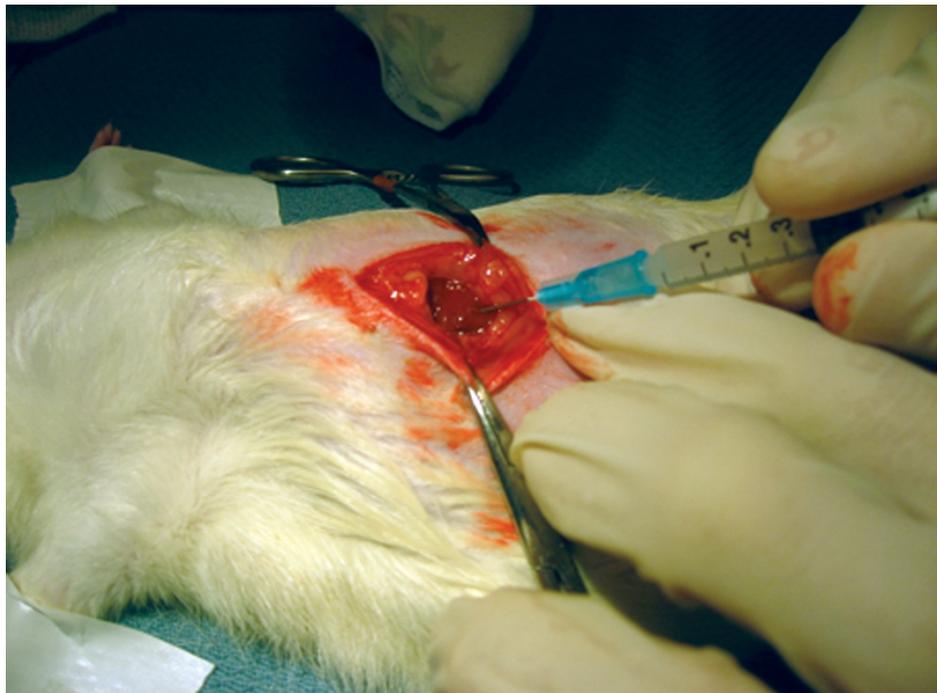
The McA-RH7777 cell line, a Morris HCC cell line derived from the Buffalo rat, was purchased from the American Type Culture Collection (Manassas, VA) and grown in a humidified incubator at 37°C, 5% CO<sub>2</sub> in Dulbecco's Modified Eagle's medium with 10% heat-inactivated fetal bovine serum sans antibiotics (Invitrogen, Carlsbad, CA). The cells were stably transfected with a retrovirus encoding click beetle red luciferase (CBRLuc) [21]. In brief, replication-incompetent retroviruses were created by transfecting Phoenix-ecotropic cells with the plasmid pMSCVpuro (Clontech Laboratories, Inc., Mountain View, CA) carrying the gene for click beetle red luciferase under beta-actin promoter control. The resultant retroviral supernatant was used to transfect Morris hepatoma McA-RH7777 cells. Reporter-expressing cells were then enriched by selection with puromycin (8 ug/mL) (Sigma, St. Louis, MO) for a minimum of five passages.

### *In vitro* bioluminescence imaging (BLI)

To compare the bioluminescent signal with cell number, the concentration of log phase McA-RH7777-CBRLuc cells, harvested eight passages after transduction, was determined by trypan exclusion assay using a hemocytometer. Cells were seeded in triplicate at a density of 1x10<sup>5</sup> cells/well into a 96-well, black wall tissue culture plate with clear bottom wells. Seven serial 1:1 dilutions of cell concentration were then performed, and the cells were incubated for four hours to allow adherence. Imaging was performed with the Xenogen In Vivo Imaging System (IVIS Spectrum; Caliper Life Sciences, Hopkinton, MA) after the addition of D-luciferin (0.15mg/mL final) (Biosynth AG, Staad, Switzerland). The charge-coupled device (CCD) camera was cooled to between -105° C and 120° C and the field of view set to 25 cm. The average radiance of uniform regions of interest at peak signal output was measured using Living Image 3.0 software (Caliper Life Sciences, Hopkinton, MA).

### Animal tumor model

Our study protocol (Protocol #26869) was approved and monitored by the Stanford Institutional Administrative Panel on Laboratory Animal Care and performed in concordance with the ethical treatment of animals. Tumor implantation was performed in six adult male Buffalo rats (300-450 gm.) (Charles River Laboratories, Wilmington, MA). The procedure was done under anesthesia with 2% isoflurane in oxygen at 2 L/min on a heated platform. A longitudinal subxiphoid incision of approximately 2 cm was made to expose the liver. Log phase, reporter-tagged HCC cells (1.0x10<sup>6</sup> McA-RH7777-CBRLuc in 100µL phosphate-buffered saline), used within 25 passages from the time of transduction, were injected under direct visualization through a 30-gauge needle into the left lateral lobe of the liver (Figure 1). The injection site was compressed for one minute with a cotton swab to prevent cell leakage out of the liver and/or bleeding into the abdominal cavity. The incision was closed with 4-0 chromic gut suture (Ethicon, Inc., Somerville, NJ). The animals were fed ad libitum and monitored daily.



**FIGURE 1: Implantation of McA-RH7777-CBRluc cells into the Buffalo rat.**

Through a subxiphoid vertical incision,  $1.0 \times 10^6$  cells in 100  $\mu$ l phosphate-buffered saline was injected under direct visualization into the left lobe.

### ***In vivo* bioluminescence imaging**

In five of the animals, BLI was performed on Day 1, Day 2, Day 4, and then every three to five days for a total of 10 time points in 28 days with the IVIS Spectrum Imaging System as previously reported [10-12]. The CCD camera was cooled to between  $-105^{\circ}$  C and  $120^{\circ}$  C and the field of view set to 25 cm. Rats were anesthetized with 2% isoflurane in oxygen at 2 L/min. Ten minutes after the rats received an intraperitoneal injection of 150 mg/kg of D-luciferin in PBS, bioluminescence images were acquired with an exposure time of 1-120 sec, medium binning, 1 f/stop, with an open filter. A region of interest was drawn around the tumor, and the bioluminescence signal was quantified as photons/sec/cm<sup>2</sup>/steradian (p/sec/cm<sup>2</sup>/sr).

### **Positron emission tomography (PET) imaging**

Serial imaging of tumor growth was performed in five of the animals on Day 1, Day 2, Day 4 and then every 3-5 days for a total of 10 time points in 28 days with a dedicated small-animal PET scanner, microPET R4 (Concorde Microsystems, Knoxville, TN). After induction and maintenance of anesthesia with 2% isoflurane in oxygen at 2 L/min, the rats were injected with 1 mCi [<sup>18</sup>F]-fluorodeoxyglucose ([<sup>18</sup>F]-FDG) via the tail vein. The rats were kept under anesthesia to minimize muscle activity. After one hour, rats were imaged for 15 minutes in the prone position with a transaxial field of view of 22 cm. A region of interest (ROI) covering the tumor was drawn. The average count rate (counts/sec per pixel) within the ROI was determined and converted to tracer activity (mCi/mL) using a standard calibration constant. Assuming a tissue density of 1 g/mL, tracer activity (mCi/g) was then divided by the injected dose (mCi) to obtain a tissue-specific uptake index (percent injected dose per gram of tissue, %ID/g). Another ROI was obtained similarly in the right lobe of the liver, and tissue uptake (%ID/g) was obtained similarly to acquire liver background signal.

Animals underwent imaging by multiple modalities on the same day under a single period of anesthetization.

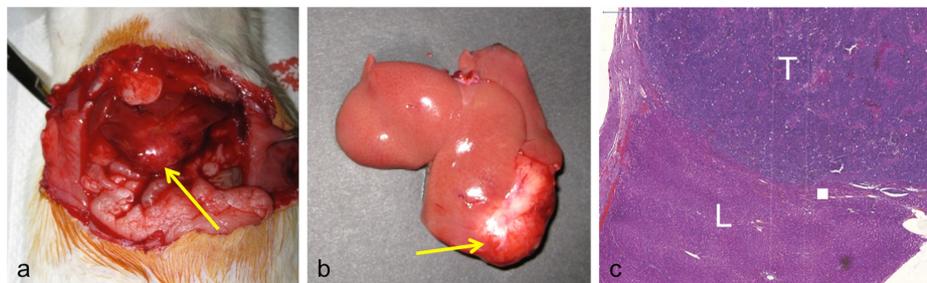
### **High-frequency ultrasound imaging**

On Day 1, Day 2, Day 4, and every 3-5 days thereafter for a total of 10 time points in 28 days post-implantation, high-frequency small animal ultrasound (US) examination of the liver was performed in five of the animals with a dedicated small-animal high-resolution imaging unit (Vevo 770; VisualSonics, Toronto, Canada). During imaging, rats remained anesthetized with 2% isoflurane in oxygen at 2 L/min on a heated stage. Real-time imaging was performed with a 40-MHz high-frequency linear transducer. Two-dimensional B-mode images were acquired in two directions (transversal and sagittal) with manual

optimization of the gain, and the maximum tumor diameter measured in millimeters was measured in millimeters using electronic calipers. Images were recorded digitally and analyzed offline using micro-US imaging software (VEVO 770; VisualSonics, Toronto, Canada).

### Euthanasia

After final imaging on Day 28, the animals were euthanized by carbon dioxide inhalation. A midline laparotomy was performed and the liver and peritoneal cavity were inspected prior to explantation of the liver. Gross pathology and hematoxylin-eosin staining from a representative liver specimen are shown in Figure 2.



**FIGURE 2: Representative gross and histopathologic specimen of Morris HCC.**

5x view reveals a sharp demarcation between the implanted tumor (upper field) and the normal liver (lower field), with evidence of hypervascularity within the tumor. 20x view demonstrates that compared with normal liver (a), the HCC (b) demonstrates poorly differentiated cells, with markedly increased mitotic activity and hyperchromatism. (c) Grossly, the tumor (arrow) is pale and well-circumscribed and extends to the liver capsule.

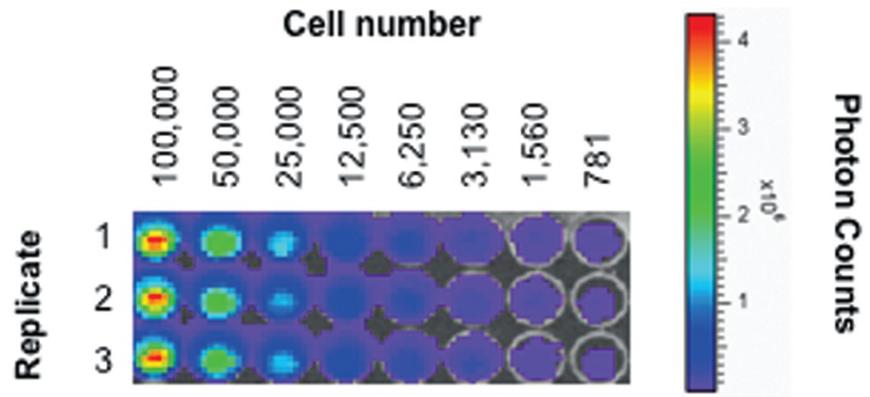
### Reproducibility of cell implantation

A subsequent 31 animals underwent implantation of stably transfected McA-RH7777 cells using methods as described above. After the initial series of six animals, the decision was made to implant tumors in the middle lobe, as we found that decreased the likelihood of the tumor adhering to the abdominal wall. All animals were imaged by PET at Day 14. The three animals, which did not have tumor detectable by PET on Day 14, were euthanized on Day 14 for evaluation. The other 28 animals underwent secondary interventions and thus were not included in the longitudinal follow-up.

## Results

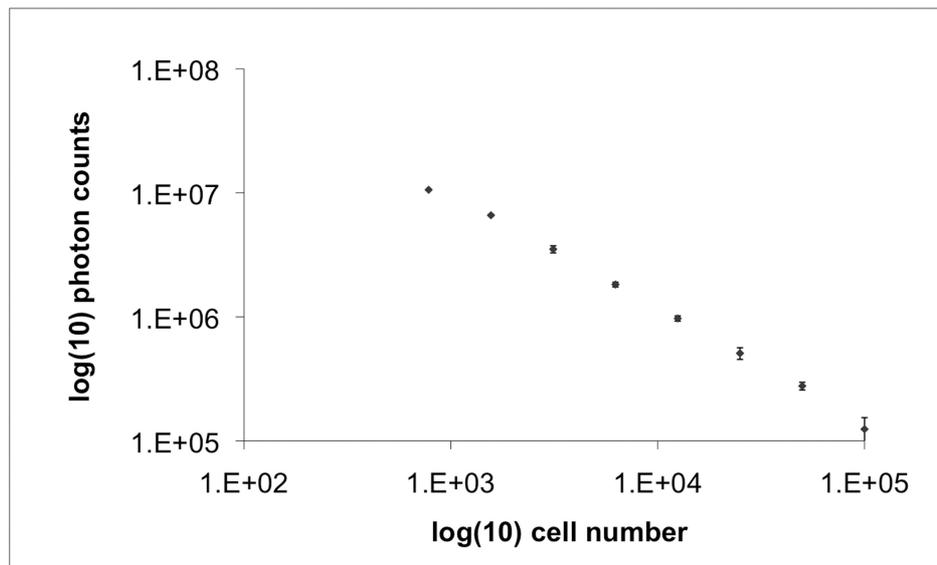
### McA-RH7777 cells demonstrate stable expression of bioluminescence genes

As shown in Figures 3-4, McA-RH7777-CBRluc cells harvested at eight passages after retroviral transfection demonstrate strong bioluminescence signal *in vitro* after the addition of D-luciferin, and a linear relationship exists between cell concentration and bioluminescence signal. Bioluminescence signal was tested at later passages and showed similar results (data not shown), confirming stable retroviral integration.



**FIGURE 3: Relationship between cell concentration and bioluminescence signal in vitro.**

Image from 96-well plate obtained with IVIS Spectrum Imaging System demonstrates bioluminescence signal from McA-RH7777-CBRLuc cells after addition of D-luciferin.

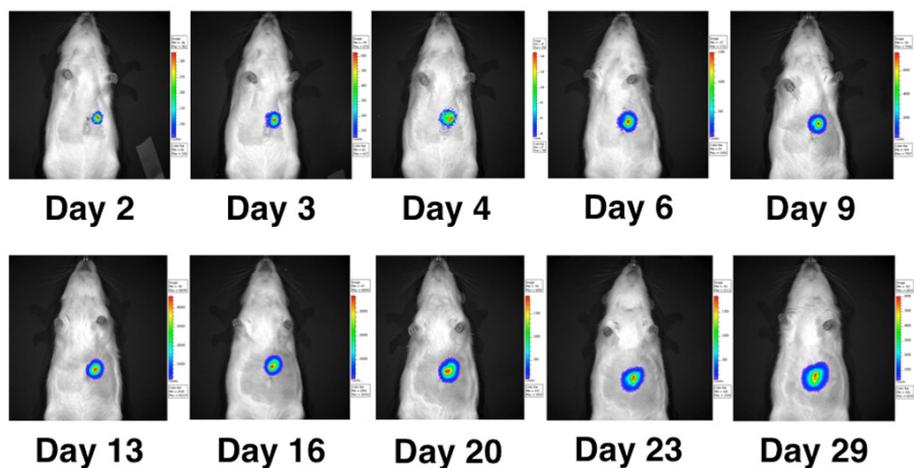


**FIGURE 4: Quantitative relationship between cell concentration and bioluminescence signal in vitro.**

The plot shows a linear relationship between cell count in vitro and bioluminescence signal. Vertical bars represent standard deviation.

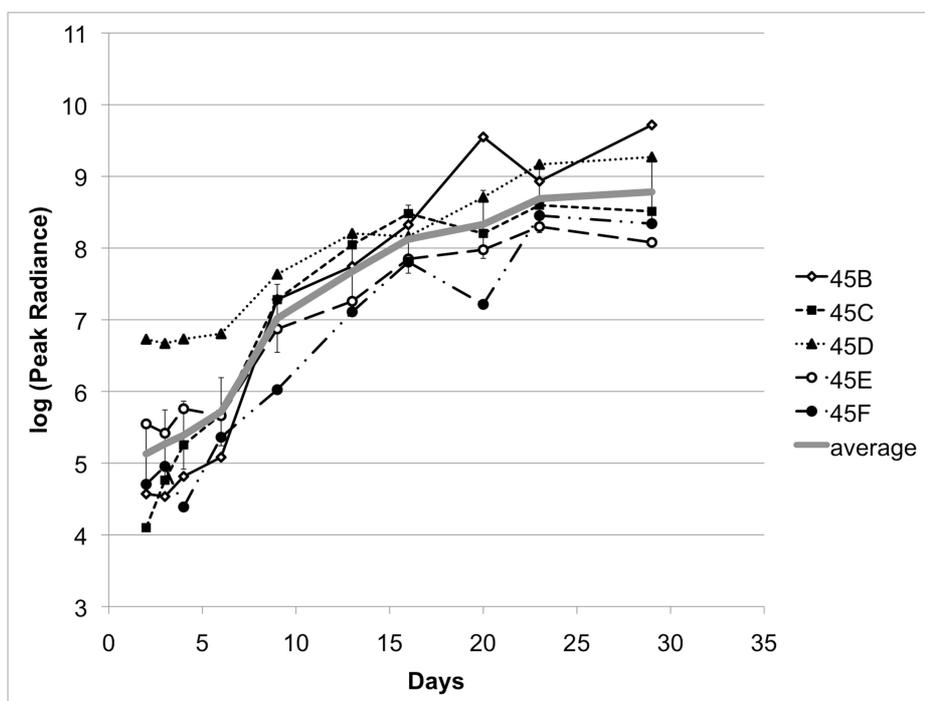
### Monitoring tumor growth *in vivo* with BLI

In the five animals that underwent serial bioluminescence imaging, tumor growth was first detected with BLI at Day 4 post-injection with a mean  $2.9 \times 10^4$  p/sec/cm<sup>2</sup>/sr (range  $4.7 \times 10^3 - 1.5 \times 10^5$ ). The BLI signal increased linearly with the tumor size before reaching a plateau on Days 21-28 (Figures 5-6).



**FIGURE 5: Serial measurements of BLI signal.**

Bioluminescence (p/sec/cm<sup>2</sup>/sr) was measured on days 2, 3, 4, 6, 9, 13, 16, 20, 23, and 29 in 5 animals.

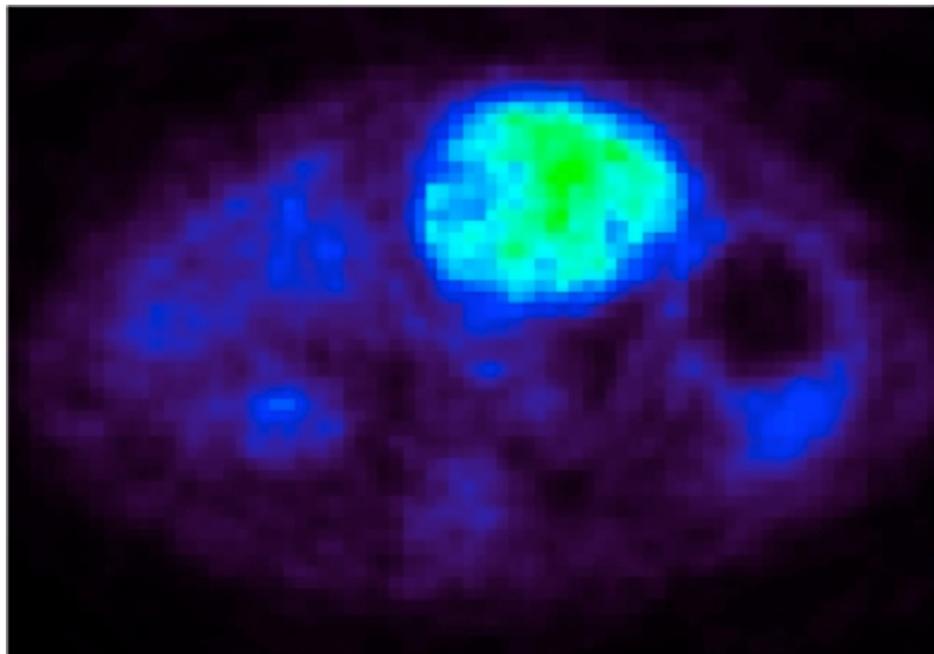


**FIGURE 6: Quantitative values of serial measurements of BLI signal.**

Bioluminescence (p/sec/cm<sup>2</sup>/sr) was measured on days 2, 3, 4, 6, 9, 13, 16, 20, 23, and 29 in 5 animals. Mean values (circles) and ranges (vertical bars) are shown.

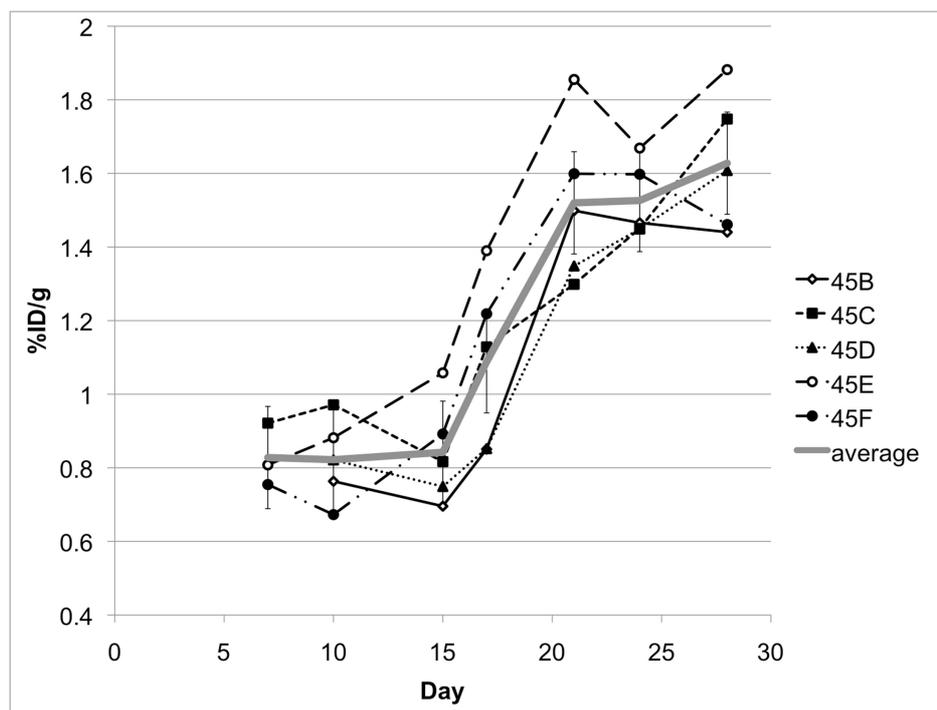
### Monitoring tumor growth with PET imaging and ultrasound

Serial PET imaging was performed at 10 time points. During PET image acquisition, the liver background signal remained constant with a mean tissue-specific uptake index of 0.04%ID/g (range 0.03-0.05). All six tumors demonstrated FDG uptake. A discrete tumor was first observed on Days 14-15 in all animals with a mean tissue-specific uptake index of 0.14%ID/g. The PET signal increased linearly with tumor growth with a maximum mean tissue-specific uptake index of 0.81%ID/g (range 0.59-0.91) on Day 28 (Figures 7-8). After Day 24, PET imaging revealed evidence of intraperitoneal tumor metastases in all animals, which was confirmed on necropsy.



**FIGURE 7: Serial measurements of PET signal.**

PET signal was measured on Days 7, 10, 15, 17, 21, 24, and 28 in 5 animals.



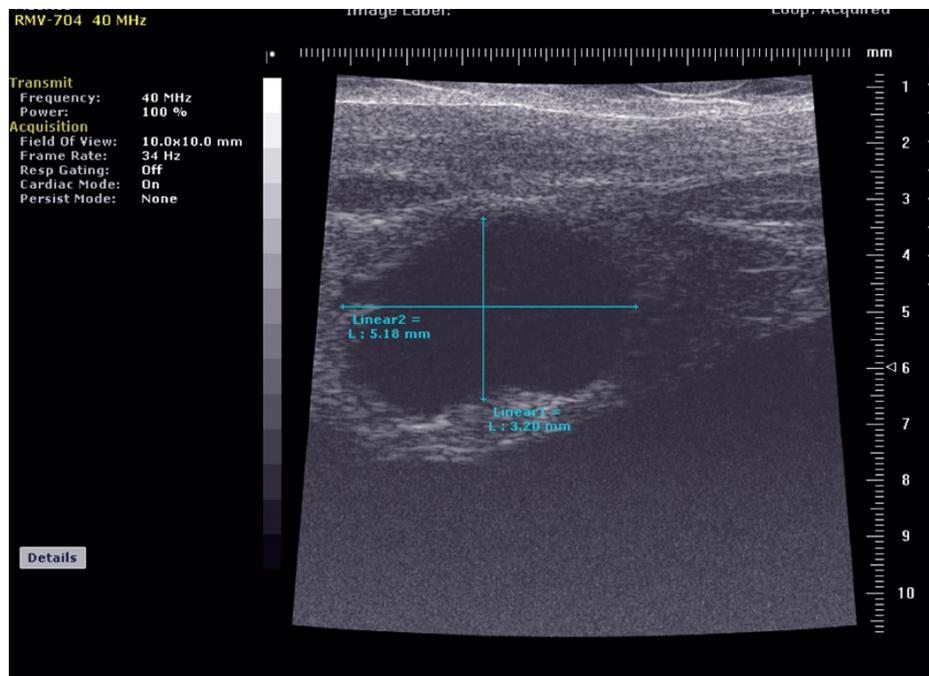
**FIGURE 8: Quantitative serial measurements of PET signal.**

PET signal (%ID/g, triangle) was measured on Days 7, 10, 15, 17, 21, 24, and 28 in 5 animals. Mean values and ranges (vertical bars) are shown.

In the 31 animals with middle lobe tumor implants, PET imaging was performed on Day 14. Three animals failed to demonstrate a hypermetabolic PET signal in the site of implantation; those animals were euthanized, and no tumor was seen on gross inspection. The mean tissue specific uptake index on Day 14 in the other 28 animals was 0.65%ID/g (range 0.35 to 0.87). The increased mean PET signal at Day 14 in these

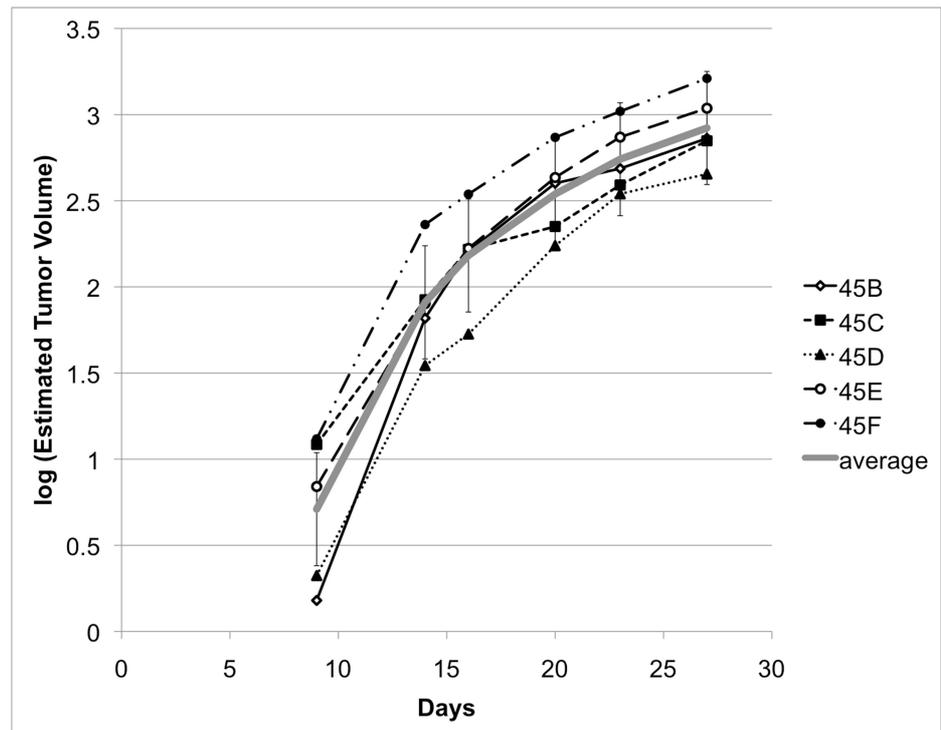
31 animals, when compared with the original six animals, is attributable to the implantation of late log phase cells (70% confluence) in the initial six animals, which would lead to slower tumor growth, versus mid-log phase cells (50% confluence) in the subsequent 31 animals. Successful tumor implantation was achieved in 34 of 37 (91.9%) of the animals.

On Days 10-12, a discrete hypochoic liver tumor was observed in all five serially imaged animals by high-frequency ultrasound with a mean maximum tumor diameter of 2.7 mm (range: 2.1-4.6). The tumors grew to a mean maximum of 11.3 mm (range 10.5-12.2) by Day 28 (Figures 9-10).



**FIGURE 9: Representative image from ultrasound imaging on Day 17 from a single animal.**

Serial estimates of tumor volume ( $\text{mm}^3$ ) were calculated from high-frequency ultrasound measurements in three dimensions, taken on Days 3, 9, 14, 16, 20, 23, and 27.



**FIGURE 10: Quantitative representation of tumor volume from ultrasound.**

Serial estimates of tumor volume ( $\text{mm}^3$ ) were calculated from high-frequency ultrasound measurements in three dimensions, taken on Days 3, 9, 14, 16, 20, 23, and 27. Graph represents log of estimated volumes. Gray line represents average of log values; vertical bars represent standard error. No appreciable tumor was seen on Day 3 for all animals.

## Discussion

Driven by the need to study the pathophysiology and treatment of HCC, numerous groups have developed animal models of HCC that range in size from the mouse to the pig. Two basic strategies have been implemented in the creation of animal models of HCC: the generation of spontaneous tumors in the liver or the implantation of tumor cells into the animal.

Spontaneous tumor models are particularly useful in investigating the pathogenesis of HCC, as the lesions created by this technique go through stages of DNA damage and dysplasia as would be expected in the natural course of the disease. These models are also more likely to have tumors developing in a background of cirrhosis or fibrosis, mirroring what is present in the majority of patients with HCC. One technique for inducing HCC is the administration of the carcinogen diethylnitrosamine (DEN) with or without phenobarbital. After receiving DEN over several weeks either through diet or injections, the subjects develop multifocal HCC. This technique is most widely used in rats [22-29] but has also been described in rabbits [30] and in pigs [31]. Other methods to generate spontaneous HCC in rats include the administration of the carcinogen *p*-dimethylaminoazobenzene [32] or implementation of a methyl-deficient [33] or choline-deficient ethionine-supplemented diet [34].

Spontaneous HCC formation without chemical intervention has also been documented in a number of animal models. A strain of rat, the Long-Evans cinnamon rat, is predisposed to hereditary hepatitis with eventual development of HCC [35]. Various transgenic mice have demonstrated a high incidence of HCC [36-40]. A high incidence of HCC formation is also seen in the Eastern American woodchuck when infected with the woodchuck hepatitis virus, which is similar to the human hepatitis B virus [41-43].

A major disadvantage of spontaneous tumor models of HCC is the long incubation time prior to tumor formation, approximately 12-16 weeks for rat carcinogen models and several months for the rabbit and pig carcinogen models. This delay is not conducive to high-throughput studies. In addition, multiple tumors develop, and the growth trajectory of the tumors is variable. In the rat spontaneous HCC models, many of the study subjects died from the natural course of the disease prior to completion of the studies. This inherent variability in the time course of tumor development would limit the investigator's ability to detect differences in the outcome after treatment.

Direct tumor implantation into animal subjects confers the advantages of rapid tumor development and control over tumor location. A facile method of tumor implantation is in the subcutaneous tissues of the animal [44-45]. However, while this technique may allow for assessment of the efficacy of intravenous therapies and direct intratumoral injections, intra-arterial delivery cannot be assessed, and the heterotopic localization may alter the tumor's behavior compared with implantation in the liver.

Consequently, orthotopic implantation of tumor into the liver has been explored by various groups. Orthotopic mouse models of HCC have been created by xenotransplantation of human HCC cells into nude mice [46-51]. While this technique has the benefit of using human HCC cells, the behavior of the tumor and the therapies may be altered by the immunocompromised state of the host. Orthotopic implantation of murine HCC into immune-competent mice has also been described [52]. The small size of the mouse, however, allows for investigation of only systemic and, possibly, direct intratumoral administration of therapy, as the vessels are too small for intra-arterial delivery.

In contrast with the mouse, the vasculature of the rat is of adequate size to permit intra-arterial delivery of therapies, in addition to intratumoral and intravenous delivery. Orthotopic rat models of HCC have been developed in Buffalo rats using McA-RH7777 or ACI rats using Morris hepatoma 3924A [16-20]. The rat's relatively small size allows for ease of handling, and the costs are substantially reduced when compared with larger animals, allowing for high-throughput studies. The small size is also advantageous in that tumors can be tracked longitudinally in a non-invasive fashion not only with PET imaging but also with BLI, which has a more limited penetration of tissue. This obviates the difficulties associated with generating and handling radiotracers.

In this paper, we describe a novel rat orthotopic HCC model that uses stably transduced tumor cells to allow longitudinal tracking with multiple non-invasive imaging modalities. While the implantation of Morris hepatoma cells has been described previously, our model is novel in the use of stable bioluminescence gene expression to allow tracking of tumor growth. The bioluminescence marker that we have selected - click beetle red luciferase - is notable in that it emits red light, which is attenuated less in deep tissues than light of shorter wavelengths [53-54].

This is also, to our knowledge, the first report demonstrating that implanted McA-RH7777 cells show a positive signal with [18]F-FDG PET imaging, providing a second means of tumor tracking. Of note, [18]F-FDG PET provides physiologic, as opposed to anatomic, imaging, and may be a better marker of tumor viability in the small animal than CT, MRI, or ultrasound. Likewise, as BLI signal can be generated only by live cells, it may also better track tumor viability than conventional anatomic imaging modalities. The ability to track change in tumor viability non-invasively is key for high-throughput evaluations of novel therapies, as it reduces the number of animals required to demonstrate statistical significance and avoids the errors inherent in assuming that all implanted animals must begin with the same tumor size when studying potential therapies. The ability to detect the tumor by ultrasound, as we found to be possible, allows the potential for percutaneous intratumoral delivery of therapies.

This model also has the advantage of rapid tumor development when compared with spontaneous models. All animals with tumor growth demonstrated PET signal in 14-15 days, and the six animals that underwent BLI demonstrated signal in four days.

A limitation of this model for intra-arterial delivery, when compared with larger animal models, is that it cannot be used for repeat intra-arterial delivery. Also, lobar delivery cannot be performed, as injection must be performed via a ligated gastroduodenal artery in order to cannulate a vessel of adequate size. In addition, unlike with spontaneous HCC models, the background liver parenchyma is non-cirrhotic, which differs from the majority of HCC patients. Lastly, it would be convenient if a serum marker for tumor growth were available. Although the ability to correlate tumor size with rat alpha-fetoprotein has been described [55], to date, we have not found a commercial test for alpha-fetoprotein that demonstrates cross-reactivity with rat alpha-fetoprotein to allow serum monitoring (data not shown).

## Conclusions

As novel pharmacologic, genetic, and combination therapies are being developed to treat HCC, investigators will need to be able to rapidly, and relatively inexpensively, test the therapies for safety and efficacy prior to large animal or human studies. With this model, high-throughput experiments can be performed using targeted routes of delivery - intra-arterial and intratumoral - in addition to intravenous delivery. The use of molecular imaging techniques in this small-animal model to track changes in tumor size provides a new platform with which investigators can advance the field of image-guided interventions in the treatment of HCC.

## Additional Information

### Disclosures

**Human subjects:** All authors have confirmed that this study did not involve human participants or tissue.

**Animal subjects:** Stanford Administrative Panel on Laboratory Animal Care Issued protocol number 26829.

**Conflicts of interest:** In compliance with the ICMJE uniform disclosure form, all authors declare the following: **Payment/services info:** All authors have declared that no financial support was received from any organization for the submitted work. **Financial relationships:** All authors have declared that they have no financial relationships at present or within the previous three years with any organizations that might have an interest in the submitted work. **Other relationships:** All authors have declared that there are no other relationships or activities that could appear to have influenced the submitted work.

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